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Roles of m⁶A RNA Methylation Modification in Cancer Stem Cells: New Opportunities for Cancer Suppression

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ABSTRACT

As a reversible post-transcriptional modification, N⁶-methyladenosine is the most common form of RNA modification in eukaryotic mRNA. Cancer stem cells (CSCs), which are a subpopulation of cells with self-renewal ability and differentiation potential, have been regarded to one of the roots of tumor occurrence, recurrence, and metastasis. Currently, numerous studies have demonstrated that m⁶A RNA modification is critically implicated in the regulation of CSCs stemness or the CSC-like traits of cancer cells. This review summarized the effects of m⁶A RNA modification-related enzymes and underlying mechanisms contributing to CSCs or cancer cell stemness, which may provide novel targets and research directions for the specifically elimination of CSCs or cancer cells with stemness.

KEYWORDS: Cancer stem cells; m⁶A RNA modification; Post-transcriptional modification; Stemness; CSC-like traits

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1. Introduction

Since the discovery of N⁶-methyladenosine (m⁶A) in 1974, m⁶A has been an unknown field due to the technical bottleneck [1]. M⁶A refers to the methylation modification process on the sixth nitrogen atom of RNA adenine catalyzed by methyltransferase, which is reversible [2]. In recent years, with the breakthrough of detection technology and the deepening of scientists' understanding of m⁶A, m⁶A-RNA modification has become numerous dynamically regulated modifications in the entire transcriptome. It has been found that m⁶A is the most common form of RNA modification in eukaryotic mRNA. There are three types of enzymes involved in the process of RNA methylation modification: Methyltransferases, also known as "writers", including methyltransferase like protein (METTL) 3, METTL14, and WT1 Associated Protein (WTAP); Demethylase is called "eraser", including Fat mass and obesity-associated protein (FTO) and alkylation repair homolog protein 5 (ALKBH5); The m⁶A recognition protein is called "reader", including the protein family of the YTH domain (YTHDFQ, YTHDF2, YTHDC1, YTHDC2, etc.) and the hnRNP protein family (hnRNPA2B1). M⁶A-RNA modification is tightly involved in transcriptional regulation and participates in almost every stage of RNA metabolism, including RNA processing, nuclear output, translation, and RNA degradation [3] (**Figure 1**).

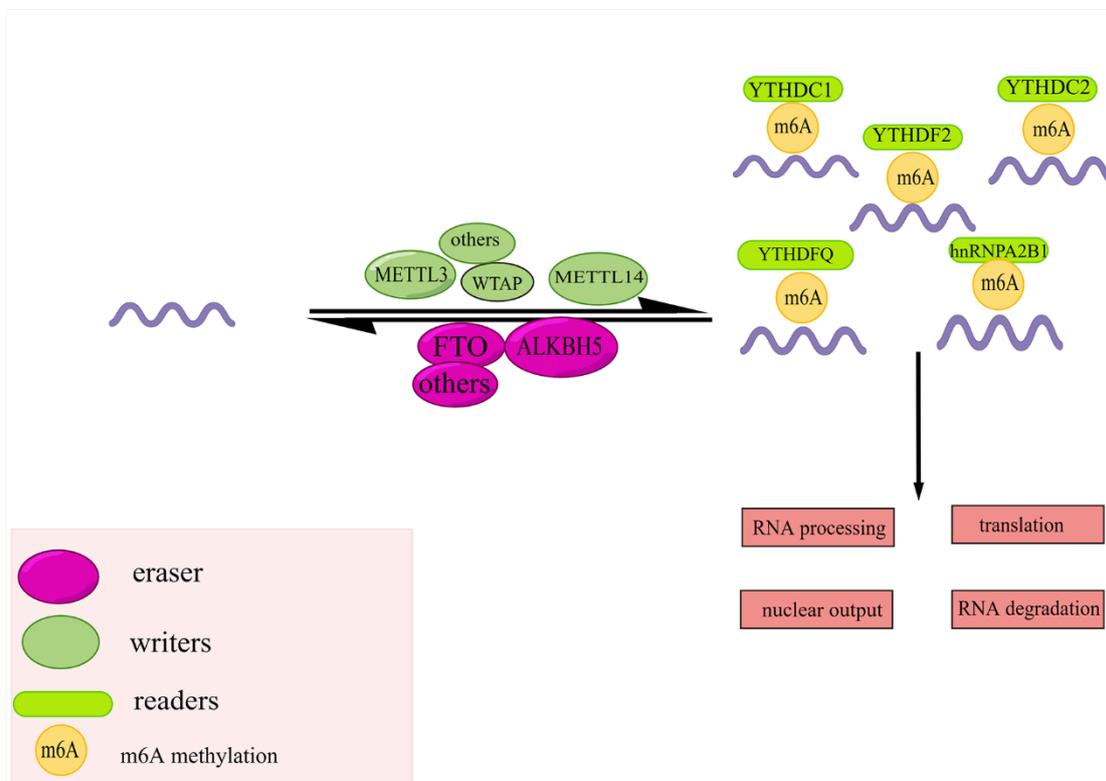


Figure 1. The process of m⁶A RNA methylation modification.

There are three types of enzymes involved in the process of RNA methylation modification.

Methyltransferases including METTL3, METTL14, and WTAP; Demethylase including FTO and ALKBH5; The m⁶A recognition protein including the protein family of the YTH domain (YTHDFQ, YTHDF2, YTHDC1, YTHDC2, etc.) and the hnRNP protein family (hnRNPA2B1). M⁶A-RNA modification is tightly involved in transcriptional regulation and participates in almost every stage of RNA metabolism, including RNA processing, nuclear output, translation, and RNA degradation.

Cancer stem cells (CSC) are a kind of cells with self-renewal ability and differentiation potential, these two specific traits of CSCs have been demonstrated to result in the tumor resistance to standard treatment methods [4]. Therefore, CSCs have long been regarded as the source of drug resistance, tumor relapse, and metastasis (**Figure 2**). Emerging evidences have revealed the critical roles of on m⁶A-RNA modification in CSCs progression [5]. This review has made a further understanding of the function of m⁶A-RNA modification, and summarized the relationship between m⁶A and CSCs and its potential applications.

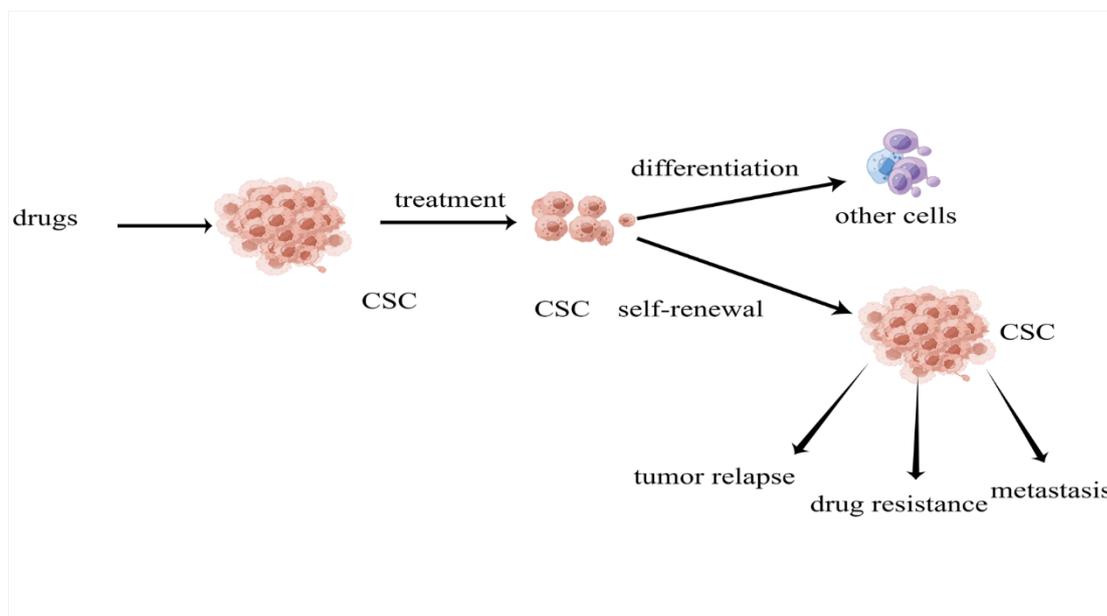


Figure 2. The functions of CSCs during tumor progression.

CSCs hold the self-renewal and differentiation ability resulting tumor relapse, drug resistance, and metastasis.

2. The constitutes of m⁶A-RNA modification and their emerging roles

Currently, more than 100 chemical modifications have been found in coding and non-coding RNA (ncRNA) [6]. M⁶A-RNA modification is the most common and abundant post-transcriptional RNA modification in eukaryotic cells. M⁶A methyltransferase and demethylase are involved in the dynamic and reversible regulation of the level of m⁶A modification. It is usually found that the m⁶A-RNA modification is concentrated

around the termination codon, which implies its role in translation control, or in the 3'-untranslated regions (3'-UTR), which affects the affinity of specific RNA binding proteins (RBPs) to their target mRNA [7]. The m⁶A modification is not random, but mainly occurs in adenine with RRACH (R: A/, H: A/C/U) conservative structure [8]. The m⁶A modification recognition protein specifically recognizes m⁶A modification and regulates RNA splicing, transport, stability and translation [9]. Although RNA methylation does not affect base pairing and gene coding, emerging studies have shown that m⁶A is widely involved in the regulation of biological processes, including stem cell renewal and differentiation, tissue development, heat shock response, tumor invasion and other processes [10, 11]. M⁶A RNA modification is involved in almost all major biological processes from normal development to disease through m⁶A methyltransferase, m⁶A demethylase, and m⁶A recognition protein.

2.1 M⁶A methyltransferase

M⁶A methyltransferase, also known as "writer", is responsible for promoting RNA methylation during the post-transcriptional modification of RNA, that is, the m⁶A process [12]. Currently, the "writer" contains three enzymes, namely METTL3, METTL14, and WTAP. Methyltransferase plays an important role in regulating biological phenomena such as biological clock, immunity, reproduction, and the occurrence and development of various diseases [13]. In the process of RNA methylation, METTL3, METTL14, and WTAP form a special complex, called WMM complex. A recent study proposed the following model: WTAP firstly binds to mRNA, and then recruits METTL3 and METTL14 complexes to catalyze the methylation of the sixth nitrogen atom on RNA adenine [14]. This WMM complex mediates methylation process and thereby regulates tissue differentiation, cell apoptosis and cell cycle, and plays critical roles in tissue formation and embryonic development (**Figure 3**).

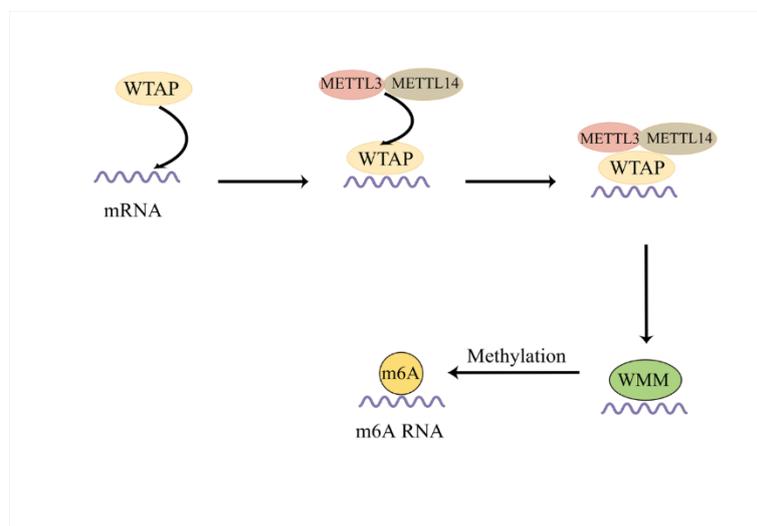


Figure 3. METTL3, METTL14, and WTAP form a special complex, called WMM complex, to mediate RNA methylation.

WTAP firstly binds to mRNA, and then recruits METTL3 and METTL14 complexes to catalyze the methylation of the sixth nitrogen atom on RNA adenine.

2.2 *M⁶A demethylase*

In 2011, the first *m⁶A* demethylase FTO was found, proving that *m⁶A* is a reversible and controllable process, which triggered a boom in *m⁶A* research [15]. In eukaryotic cells, “erasers” include FTO and ALKBH5, both of which belong to AlkB α -lutaric acid dependent dioxygenase [16]. Before 2011, most studies focused on the relationship between FTO and obesity and eating [17]. Thus, FTO is also called obesity gene. With the function of FTO as a demethylase widely known, research on FTO-mediated *m⁶A*-RNA regulation and thus affecting the occurrence and progress of diseases (especially cancer) began to boom [18]. Knockout of FTO or ALKBH5 in human cells resulted in an overall upregulation of *m⁶A* levels. FTO contains a special carboxyl terminal domain and N-terminal Alk like domain [19]. These structures can identify the common sequence ACU or RRACU in the target sequence. FTO catalyzes the oxidation of *m⁶A* to form N⁶-hydroxymethyl adenine and N⁶-aldehyde adenine, and specifically catalyzes the decomposition of intermediates into adenosine and formaldehyde in a concentration dependent manner to achieve *m⁶A* demethylation. FTO targets multiple RNA substrates and has different functions for different tissues and biological systems. Another RNA demethylase, ALKBH5, widely exists in human tissues and is mainly localized in the nucleus, especially abundant in mouse testes [20]. It is essential for spermatogenesis and mouse reproduction, and functionally contributes to the normal splicing of mRNA and the formation of longer 3'-UTR mRNA. In addition, ALKBH5 can promote the demethylation of *m⁶A* and affect the metabolism, stability, nuclear output, splicing and translation efficiency of mRNA [21]. Knockout or overexpression of ALKBH5 will affect the expression level of mRNA methylation and cause many diseases. Downregulation of ALKBH5 can accelerate the nuclear output of mRNA and is closely related to reproductive system diseases and various cancers through *m⁶A*-dependent modification (**Figure 4**).

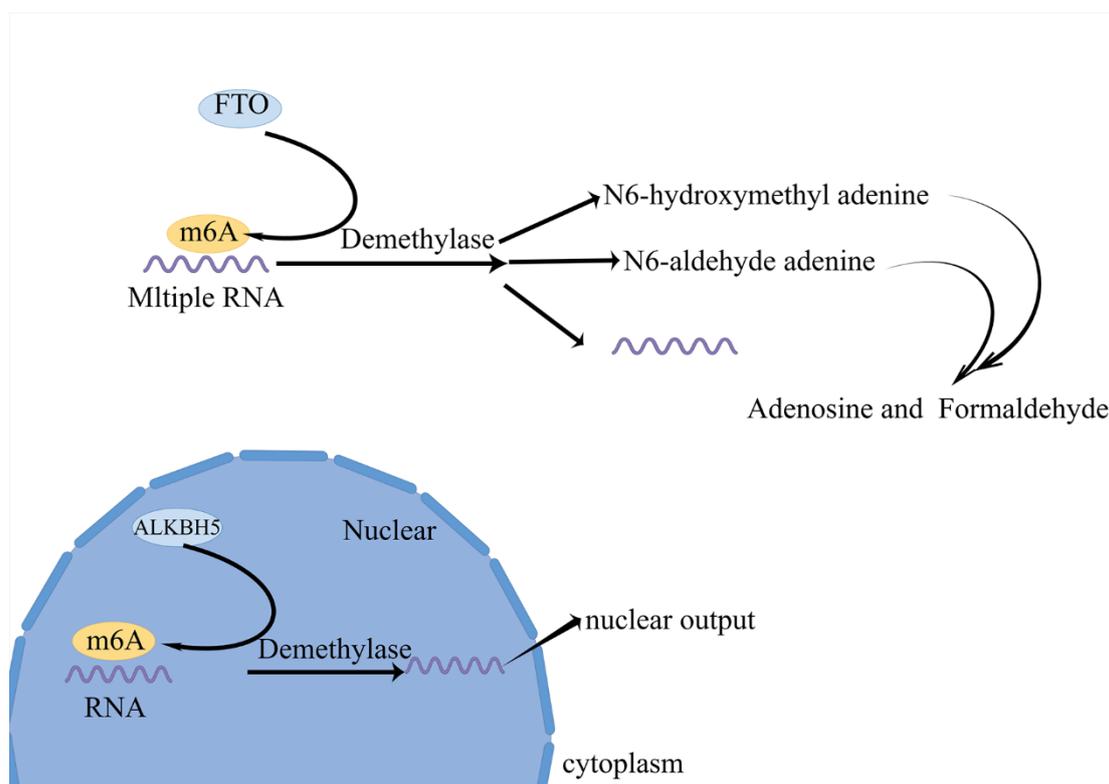


Figure 4. The process of M⁶A demethylase-mediated RNA demethylation. FTO catalyzes the oxidation of m⁶A to form N6-hydroxymethyl adenine and N6-aldehyde adenine, and specifically catalyzes the decomposition of intermediates into adenosine and formaldehyde in a concentration dependent manner to achieve m⁶A demethylation. ALKBH5 is mainly localized in the nucleus and functionally promotes the demethylation of m⁶A and affect the metabolism, stability, nuclear output, splicing and translation efficiency of mRNA.

2.3 M⁶A recognition protein

The m⁶A recognition protein is called "reader", which regulates the interaction between RNA and protein to enable the m⁶A-modified RNA to perform specific biological functions [22]. It mainly recognizes m⁶A modification in RNA and regulates downstream molecular mechanisms. "Readers" mainly include: YTHDFs and YTHDCs subtypes; HNRNPs, IF2BPs et al [23]. Methylated-reading proteins can selectively recognize and bind to m⁶A-modified mRNA to regulate downstream pathways. Functionally, YTHDF2 can recruit mRNA containing m⁶A modification to the mRNA decay point, and reduce the stability of RNA through transcription complex subunit 4 (CCR4-NOT)-mediated adenosine acidification, thus mediating the degradation of transcripts [24]. YTHDF1 interacts with translation initiation factors to promote the translation of target mRNA [25]. YTHDF1 interacts with YTHDF3 to enhance the translation effect and affect YTHDF2-mediated RNA degradation [26]. It has been shown that YTHDF1-3 interacts with each other to

jointly regulate RNA metabolism. IGF2BP1 -3 increases the stability of mRNA containing m⁶A modification to protect RNA in cells [27]. In the mouse lung cancer model, IGF2BP1 can cooperate with kirsten rat sarcoma viral oncogene (KRAS) to promote tumor progression [28]. In animal models, YTHDF1 can affect the function of tumor antigen presenting CD8⁺T cells in mice [29]. Additionally, YTHDF1 may be a potential therapeutic target to enhance the efficacy of PD-L1 [30]. YTHDF2 affects all aspects of RNA metabolism and plays an important role in many biological processes, such as migration, invasion, metastasis, proliferation, apoptosis, cell cycle, cell vitality, cell adhesion, differentiation and inflammation of human cancer [31]. The consumption of all three YTHDF analogues can promote the differentiation of leukemia cells. Notably, all YTHDF proteins have the same m⁶A binding site on mRNA (**Figure 5**).

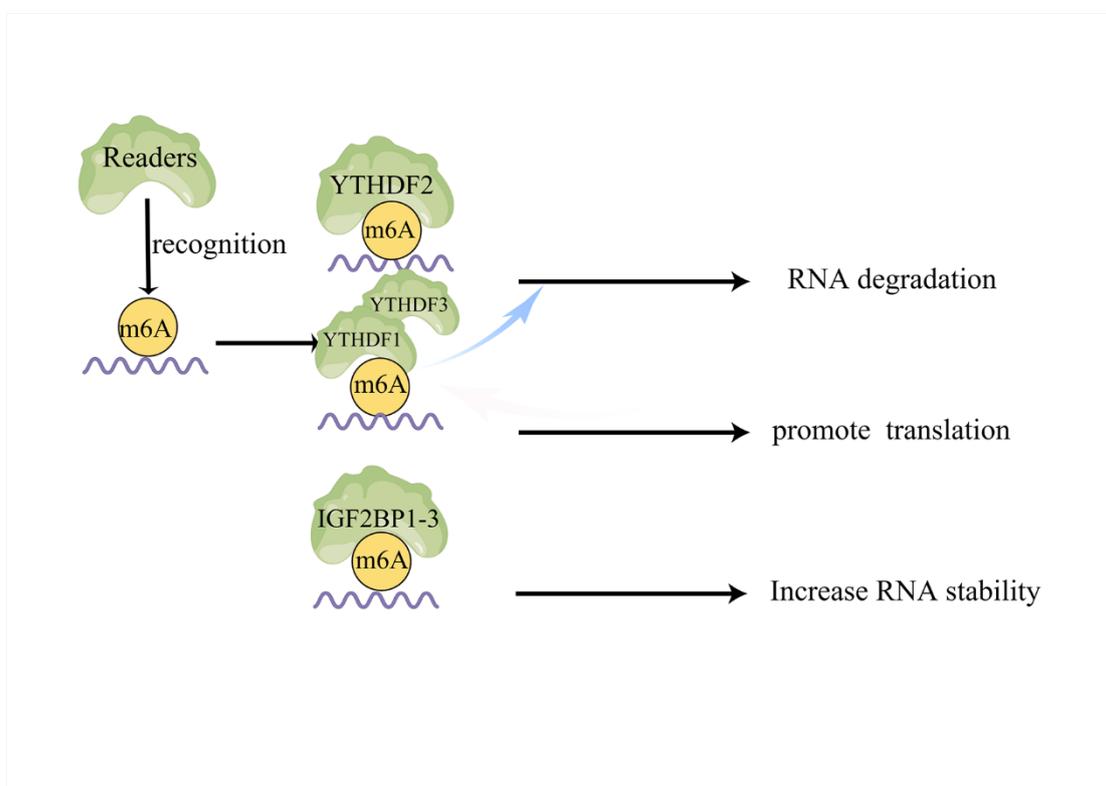


Figure 5. The process of m⁶A RNA methylation mediated by M⁶A recognition protein. YTHDF1 interacts with translation initiation factors or YTHDF3 to promote the translation of target mRNA and affect YTHDF2-mediated RNA degradation. Additionally, YTHDF1-3 interacts with each other to jointly regulate RNA metabolism. Furthermore, IGF2BP1 -3 increases the stability of mRNA containing m⁶A modification to protect RNA in cells.

Through the action of the above three enzymes, m⁶A becomes a dynamic and reversible process, and participates in RNA metabolism through methylation, affecting upstream and downstream genes.

3. The effects of m⁶A-RNA modification on CSC progression

CSCs are a group of cells with self-renewal and multi-directional differentiation potential. It is generally believed that the basic characteristics of CSCs are self-replication and multi-lineage differentiation and cloning *in vitro* and *in vivo*, which can produce cancer cells. Therefore, CSCs not only play an important role in cancer progression and maintaining the homeostasis of cancer cells, but also have broad application prospects in the treatment of different kinds of tumors. Experiments showed that CSCs have higher proliferative capacity than ordinary cancer cells. They can divide and proliferate continuously for many times, which is the primary condition to maintain the stability of cancer cell characteristics. Additionally, the differentiation potential, that is, CSCs derived from a single cell can produce and differentiate into cancer cells. Furthermore, CSCs have been found to have immunomodulatory activity as CSCs have been shown to exhibit the immunotherapeutic resistance. According to these characteristics, CSCs play an irreplaceable role in the treatment of malignant tumors as our previous studies have shown targeting CSCs can significantly suppress tumor migration, invasion, and drug resistance [32-34]. Epigenetics plays an important role in maintaining the above biological characteristics of CSCs, which refers to the phenomenon that the DNA sequence has not changed but the traits have heritable changes. In recent years, with the rapid development of high-throughput sequencing technology, epigenetic modification, especially epigenetic post-transcriptional modification, has gradually become one of the important hotspots in disease occurrence and development. Among them, m⁶A is the most abundant type of apparent post-transcriptional modification in eukaryotic mRNA. The methylation modification of m⁶A targets downstream molecules through its corresponding methylase molecules, and is widely involved in CSC proliferation and self-renewal, targeted differentiation, immune regulation and other biological processes [35]. Recent studies have shown that the occurrence and development of many malignant tumors involve the modification of corresponding CSCs by m⁶A RNA modification. Through the regulation of corresponding m⁶A-RNA modification, including the regulation of RNA splicing, exonuclear transport, translation and stability, cells can respond quickly to external stimuli, which can achieve the treatment of malignant tumors and remission of cancers. With the continuous development of epigenetics and the in-depth study of tumor treatment, m⁶A RNA modification plays a very important role in regulating the occurrence and development of CSCs-related malignant tumors.

3.1 The effects of m⁶A methyltransferase on CSC progression

METTL3 can regulate mouse embryonic stem-cell heterochromatin, the integrity of which is critical for

silencing retroviral elements and for mammalian development [36]. Since normal stem cells and CSCs share numerous common traits during their progression [37], it has been shown that knockdown of METTL3 or METTL14, the key components of the RNA methyltransferase complex, dramatically promotes human glioma stem cell growth, self-renewal, and tumorigenesis [38]. And an integrated analysis of m⁶A-RIP (RNA immunoprecipitation) and total RNA-Seq of METTL3-silenced glioma stem cells identified that m⁶A modification in glioma stem cells is principally carried out by METTL3 [39]. Consistently, Visvanathan et al. found that silencing METTL3 in glioma stem-like cells can inhibit the growth of intracranial glioma *in situ* and prolong the survival period of mice [40]; Yu-Zhou Chang et al. demonstrated that METTL3 promotes the stem cell maintenance and thus facilitates the malignant progression of glioma through the upregulation of metastasis - associated lung adenocarcinoma tran 1 (MALAT1) expression by enhancing its stability via m⁶A modification [41]. Meanwhile, silencing METTL3 could enhance the sensitivity of γ radiation in glioma stem cells. Additionally, the critical roles of METTL3 in the progression of other types of CSCs also have been largely revealed, for example, Ting Li et al. found that METTL3 significantly inhibited cell self-renewal, the frequency and migration of CSCs, and the occurrence and metastasis of colorectal cancer by targeting IF2BP2 *in vitro* and *in vivo* [42]; A recent study confirmed that METTL3-mediated m⁶A-RNA modification is necessary to activate the TEK-VEF -A-mediated tumor progression and angiogenesis in bladder CSCs [43]. Similarly, AO et al. proved that m⁶A-RNA modification played a key role in self-renewal and tumorigenicity of bladder CSCs through the METTL3-AFF4-SOX2/MYC signal axis, and promoted the progress of bladder CSCs [44]; METTL3 can also promote the stemness and malignant progression of breast cancer [45], kidney cancer [46], and gastric cancer [47]. Furthermore, METTL14, another key component of the RNA methyltransferase, is indicated to be implicated in CSC progression, like WEN et al. found that METTL14, is highly expressed in normal hematopoietic stem/progenitor cells (HSPCs) and acute myeloid leukemia (AML) cells carrying t(11q23), t(15;17), or t(8;21), and METTL14 is required for development and maintenance of AML and self-renewal of leukemia stem/initiation cells through by regulating its mRNA targets (e.g., MYB and MYC) through m⁶A modification [48]; Zhenchuan Liu et al. demonstrated that METTL14 expression was downregulated in esophageal squamous cell carcinoma, suppressed TRIB2 expression via miR-99a-5p-mediated degradation of TRIB2 mRNA by targeting its 3'-UTR, this is responsible for the enhanced CSC properties and radiotherapeutic resistance [49]; And METTL14 knockout promotes the proliferation, self-renewal, metastasis and tumor initiating capacity of bladder CSCs via regulating the

stability of Notch1 mRNA through m⁶A modification [50]. Notably, hexavalent chromium [Cr(VI)] is a common environmental carcinogen causing lung cancer in humans, a recent study indicated that and chronic Cr(VI) exposure could alter cellular epitranscriptome by increasing the m⁶A RNA modification via upregulating METTL3 expression, which plays an important role in Cr(VI)-induced cell transformation, CSC-like property, and tumorigenesis [51]. Besides, WTAP-mediated m⁶A methylation of Bcl-2 mRNA is shown to be necessary for recombinant neuropilin 1 (NRP1)-induced stemness and radiotherapeutic resistance in breast cancer [52].

3.2 The effects of m⁶A demethylase on CSC progression

Shen et al. confirmed that targeting m⁶A-RNA demethylase ALKBH5 can effectively inhibit the development and maintenance of AML, inhibit the self-renewal of leukemia stem cells, and retain normal hematopoietic function, highlighting the potential of targeting the LKBH5-ACC3 axis to treat leukemia [53]. Zhang et al. revealed that the increase of ALKBH5 in glioma stem cell-like cells promoted the self-renewal and tumorigenesis by regulating the ALKBH5-FOXO1 pathway [54]. In consistent, Kowalski et al. proved that the high expression of ALKBH5 increased the radiation resistance of glioma stem cells by regulating homologous recombination (HR). Interestingly, they also found that ALKBH5 promoted the invasion of glioblastoma by promoting the invasion of glioblastoma [55]. Additionally, the previous studies have shown that hypoxia induces the phenotype of breast CSCs by promoting HIF-dependent and ALKBH5-mediated Nanog mRNA demethylation [56]; Yu et al. have demonstrated that ALKBH5 can promote the tumorigenicity of multiple myeloma by increasing the proportion of side population (SP) cells and CD138-/CD34 myeloma stem cells by activating the Hippo signal pathway related to CSCs and promoting the expression of multifunctional factors NANO, SOX2 and OCT4 [57]. Furthermore, ALKBH5 was found to be highly expressed in CSCs derived from non-small cell lung cancer (NSCLC) and could suppress lung cancer progression by regulating epithelial-mesenchymal transition (EMT) and stemness via modulating p53 gene transcriptional activity [58]. These data indicate that ALKBH5 plays a key role in the maintenance of multiple myeloma stem cells.

FTO, another important m⁶A demethylase, also has been found to be involved in CSC progression, such as Huang et al. found that FTO enhanced the second messenger cAMP signal, inhibited the self-renewal of ovarian CSCs, and thus suppressed the occurrence of tumors by exerting the activity of demethylase [59]. Similarly, Sébastien Relier et al. demonstrated that FTO impedes CSC abilities in colorectal cancer through its

N⁶,2'-O-dimethyladenosine (m⁶A_m) demethylase activity [60]. In addition, FTO was found to be highly expressed in esophageal cancer stem-like cells, and that its level was also substantially increased in esophageal cancer tissues, which was closely correlated with a poor prognosis in esophageal cancer patients; Functional experiments indicated that FTO knockdown significantly suppressed the proliferation, invasion, stemness, and tumorigenicity of esophageal cancer cells via promoting the formation of lipid droplets in esophageal cancer cells by enhancing HSD17B11 expression [61].

3.3 The effects of M⁶A recognition protein on CSC progression

As one of distinct family members of m(6)A readers, IF2BP1 promotes the stability of MAT5 mRNA by upregulating the m⁶A-RNA modification of MAT5 mRNA, thus promoting the formation of the phenotype of liver CSCs [45]. Additionally, IF2BP1 can bind to other transcripts to regulate CSC activity, such as Irina A Elcheva et al. reported that genetic or chemical inhibition of IF2BP1 decreases leukemia cells tumorigenicity and sensitizes leukemia cells to chemotherapeutic drugs through critical regulators of self-renewal (HOXB4, MYB, ALDH1A1) [62]. IF2BP1 can also bind to and stabilize m⁶A-modified IQAP3 transcript, which is an important stem cell factor in rapidly proliferating isthmus stem cells in the stomach, to sustain stem cell potential in cancer [63].

As another “readers”, YTHDF2 can target the YTHDF2-MYC-IDFBP3 axis to link RNA endonucleating transcriptome modification and maintain the expression of oncogenes, thus promoting the growth of glioma stem cells, this indicates that YTHDF2 is a potential target in glioblastoma [64]. Similarly, Zhang et al. confirmed that YTHDF2 can promote the phenotype of liver CSCs and tumor metastasis by promoting the m⁶A-RNA modification of OCT4 mRNA, leading to the enhanced expression of OCT4 protein, which is also a critical stemness regulatory master [65]. In addition, targeting YTHDFs can also regulate CSC activity or tumor cell stemness, like TRIM29, as an oncogene, promotes the stem cell-like phenotype of cisplatin-resistant ovarian cancer cells in a m⁶A-YTHDF1 dependent manner [66]; Targeting YTHDF2 can selectively compromise CSCs in AML with enhancing hematopoietic stem cells activity [67]. Furthermore, heterogeneous nuclear ribonucleoprotein A2B1 (hnRNPA2B1), which is a known m⁶A “reader” and reported to implicated in lung adenocarcinoma progression via reading the m⁶A site on primary microRNA-106b (pri-miR-106b) to facilitate the maturing of miR-106b-5p, thus activating the Wnt/ β -catenin signaling to aggravate stemness [68]; In melanoma stem cells, Mengqi Chu et al. also revealed that hnRNPA2B1 level was significantly upregulated in melanoma stem cells compared with non-stem cells and facilitated the

tumorigenesis by affecting the splicing of TPPP3, EIF3H, DOCK2, DAPK1, RNF128, and SYT7 [69]. These above results indicate that m⁶A-related enzymes are inextricably linked with CSCs by mediating post-transcriptional m⁶A modification of RNA, and m⁶A regulatory factors hold the potential as therapeutic targets.

4. The application prospect of targeting m⁶A-RNA modification in CSCs-targeted therapy

Targeting FTO has been demonstrated to hold promising therapeutic significance via suppressing tumor growth, potentiating immunotherapy, and attenuating drug resistance [18], for example, Rui Su et al. reported two potent small-molecule FTO inhibitors (FB23 and FB23-2) that exhibit strong anti-tumor effects in multiple types of cancers by dramatically attenuating leukemia stem cell self-renewal and reprogram immune response [70, 71]; Sarah Huff et al. further demonstrated that described the structure-based design, synthesis, and biochemical evaluation of a new class of FTO inhibitors (FTO-02 and FTO-04), which could attenuate neurosphere formation in patient-derived glioblastoma stem cells without affecting the growth of healthy neural stem cell-derived neurospheres [72]. Moreover, Kunxia Cao et al. revealed that FTO inhibitor-loaded SH⁻-bioimprinted nanocomposites (NPIPP12MA) can selectively target leukemia blasts, especially leukemia stem cells, and induce ferroptosis by disrupting intracellular redox status [73]. In addition, FTO makes leukemia cells sensitive to the cytotoxicity of T cells and overcomes the immune escape induced by HMA [71]. In the mouse acute myeloid leukemia model, 50 nmol of CS1 can almost completely inhibit the regeneration of leukemia cells, highlighting the strong role of FTO inhibitors in inhibiting the self-renewal of cancer cells. Interestingly, using FTO inhibitors as cancer drugs has many advantages. FTO inhibitors can prevent or treat obesity and overweight concomitantly. FTO inhibitors R-2H, MA, FB23 and FB23-2 have been proved to inhibit the activity of FTO or the process of FTO-mediated demethylation, and thus have anti-tumor effects [16, 74]. Two compounds were identified in a high-throughput virtual screening library of 144000 pre-selected compounds, which inhibited the proliferation of three leukemia cell lines [75]. Eliza Yankova et al. carried out high-throughput screening on 250000 different drug-like compounds and finally developed the METTL3 small molecule inhibitor STM2457 [76]. They further proved its inhibition on leukemia through *in vitro* and *in vivo* experiments. Similarly, a recent study also reported two novel FTO inhibitors using virtual screening, structural optimization, and bioassay, namely 18077 and 18097 exhibiting the activity of suppressing breast cancer [77]. Furthermore, some clinically-approved drugs or natural compounds also have been indicated to improve chemoresistance and CSC progression via targeting m⁶A

RNA modification, such as omeprazole [78], Saikosaponin D [79], Simvastatin [80], and Berberine [81]. Although m⁶A-related enzyme inhibitors are currently in the experimental stage, the development of inhibitors and the success of various experiments undoubtedly suggest a new way to treat cancer. It is believed that more m⁶A-related enzyme inhibitors can be used in tumor research and treatment in the near future, and we will be closer to cancer cure.

5. Discussion and conclusion

M⁶A refers to the methylation modification process on the sixth nitrogen atom of RNA adenine catalyzed by methyltransferase, which is reversible. M⁶A is involved in many processes such as tumorigenesis, development and drug resistance, and plays a role in promoting most tumors. CSCs are cells with self-renewal ability and differentiation potential, which play a key role in tumor drug resistance, recurrence and distant metastasis. In recent years, the research on the influence of m⁶A modification and abnormal expression of m⁶A regulatory protein on the stemness of CSCs is increasing. M⁶A regulatory protein can affect various CSCs-related cancers by regulating the epitope transcriptome of cancer, thereby promoting or inhibiting the CSCs stemness, drug and radiotherapeutic resistance. This suggests that m⁶A is closely related to the occurrence and development of cancer, the generation of drug resistance and prognosis. Although some studies have shown that m⁶A-related enzymes regulate the stemness of a variety of CSCs by mediating, inhibiting or reading m⁶A, targeting numerous signal pathways, the specific regulatory mechanisms are still fragmentary, which requires in-depth research. Currently, there is no detection and sequencing technology for RNA modification types, and many theoretical and basic technical problems need to be solved. Improving the RNA epigenetic regulation theory is the research basis for further analyzing epigenetics and precise treatment of diseases. Meanwhile, m⁶A level in CSCs can be used to predict cancer risk, achieve early diagnosis, predict patient prognosis, and provide new treatment methods, which has practical significance.

6. Competing interests

The authors declare that they have no competing interests.

7. Acknowledgements

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References

1. Bangerter BW, Modestas Z: Proton magnetic resonance study of complex formation between N6-methyladenosine and polyuridylic acid. *Biopolymers* 1974, 13(3):567-575.
2. Reichel M, Köster T, Staiger D: Marking RNA: m6A writers, readers, and functions in Arabidopsis. *Journal of molecular cell biology* 2019, 11(10):899-910.
3. Oerum S, Meynier V, Catala M, Tisné C: A comprehensive review of m6A/m6Am RNA methyltransferase structures. *Nucleic acids research* 2021, 49(13):7239-7255.
4. Battle E, Clevers H: Cancer stem cells revisited. *Nature medicine* 2017, 23(10):1124-1134.
5. Ma Z, Ji J: N6-methyladenosine (m6A) RNA modification in cancer stem cells. *Stem Cells Dev* 2020.
6. Zhao LY, Song J, Liu Y, Song CX, Yi C: Mapping the epigenetic modifications of DNA and RNA. *Protein & cell* 2020, 11(11):792-808.
7. Selmi T, Hussain S, Dietmann S, Heiß M, Borland K, Flad S, Carter JM, Dennison R, Huang YL, Kellner S *et al*: Sequence- and structure-specific cytosine-5 mRNA methylation by NSUN6. *Nucleic acids research* 2021, 49(2):1006-1022.
8. Lu Z, Liu J, Yuan C, Jin M, Quan K, Chu M, Wei C: m(6)A mRNA methylation analysis provides novel insights into heat stress responses in the liver tissue of sheep. *enomics* 2021, 113(1 Pt 2):484-492.
9. Huang H, Weng H, Chen J: m(6)A Modification in Coding and Non-coding RNAs: Roles and Therapeutic Implications in Cancer. *Cancer cell* 2020, 37(3):270-288.
10. Wang P, Feng M, Han , Yin R, Li Y, Yao S, Lu P, Wang Y, Zhang H: RNA m(6)A Modification Plays a Key Role in Maintaining Stem Cell Function in Normal and Malignant Hematopoiesis. *Front Cell Dev Biol* 2021, 9:710964.
11. Batista PJ, Molinie B, Wang J, Qu K, Zhang J, Li L, Bouley DM, Lujan E, Haddad B, Daneshvar K *et al*: m(6)A RNA modification controls cell fate transition in mammalian embryonic stem cells. *Cell stem cell* 2014, 15(6):707-719.
12. Dominissini D, Moshitch-Moshkovitz S, Schwartz S, Salmon-Divon M, Ungar L, Osenberg S, Cesarkas K, Jacob-Hirsch J, Amariglio N, Kupiec M *et al*: Topology of the human and mouse m6A RNA methylomes revealed by m6A-seq. *Nature* 2012, 485(7397):201-206.
13. Zhang H, Shi X, Huang T, Zhao X, Chen W, u N, Zhang R : Dynamic landscape and evolution of m6A methylation in human. *Nucleic acids research* 2020, 48(11):6251-6264.
14. u Z, Du Y, Zhao X, Wang C: Diagnostic, Therapeutic, and Prognostic Value of the m(6)A Writer Complex in Hepatocellular Carcinoma. *Front Cell Dev Biol* 2022, 10:822011.
15. Jia , Fu Y, Zhao X, Dai Q, Zheng , Yang Y, Yi C, Lindahl T, Pan T, Yang Y *et al*: N6-methyladenosine in nuclear RNA is a major substrate of the obesity-associated FTO. *Nat Chem Biol* 2011, 7(12):885-887.
16. Huang Y, Yan J, Li Q, Li J, ong S, Zhou H, an J, Jiang H, Jia F, Luo C *et al*: Meclofenamic acid selectively inhibits FTO demethylation of m6A over ALKBH5. *Nucleic acids research* 2015, 43(1):373-384.
17. Loos RJ, Yeo S: The bigger picture of FTO: the first WAS -identified obesity gene. *Nature reviews Endocrinology* 2014, 10(1):51-61.
18. Li Y, Su R, Deng X, Chen Y, Chen J: FTO in cancer: functions, molecular mechanisms, and therapeutic implications. *Trends in cancer* 2022, 8(7):598-614.
19. Marcinkowski M, Pilżys T, arbic z D, Piwowarski J, Mielecki D, Nowaczyk , Taube M, ielnik M: Effect of Posttranslational Modifications on the Structure and Activity of FTO Demethylase. *International journal of molecular sciences* 2021, 22(9).
20. Wang J, Wang J, u Q, Ma Y, Yang Y, Zhu J, Zhang Q: The biological function of m6A demethylase

- ALKBH5 and its role in human disease. *Cancer cell international* 2020, 20:347.
21. Qu J, Yan H, Hou Y, Cao W, Liu Y, Zhang E, He J, Cai Z: RNA demethylase ALKBH5 in cancer: from mechanisms to therapeutic potential. *Journal of hematology & oncology* 2022, 15(1):8.
 22. Nair L, Zhang W, Laffleur B, Jha MK, Lim J, Lee H, Wu L, Alvarez NS, Liu ZP, Munteanu EL *et al*: Mechanism of noncoding RNA-associated N(6)-methyladenosine recognition by an RNA processing complex during IgH DNA recombination. *Molecular cell* 2021, 81(19):3949-3964.e3947.
 23. Shi H, Wei J, He C: Where, When, and How: Context-Dependent Functions of RNA Methylation Writers, Readers, and Erasers. *Molecular cell* 2019, 74(4):640-650.
 24. Du H, Zhao Y, He J, Zhang Y, Xi H, Liu M, Ma J, Wu L: YTHDF2 destabilizes m(6)A-containing RNA through direct recruitment of the CCR4-NOT deadenylase complex. *Nature communications* 2016, 7:12626.
 25. Liu T, Wei Q, Jin J, Luo Q, Liu Y, Yang Y, Cheng C, Li L, Pi J, Si Y *et al*: The m6A reader YTHDF1 promotes ovarian cancer progression via augmenting EIF3C translation. *Nucleic acids research* 2020, 48(7):3816-3831.
 26. Zaccara S, Jaffrey SR: A Unified Model for the Function of YTHDF Proteins in Regulating m(6)A-Modified mRNA. *Cell* 2020, 181(7):1582-1595.e1518.
 27. Lan Q, Liu PY, Bell JL, Wang JY, Hüttelmaier S: The Emerging Roles of RNA m(6)A Methylation and Demethylation as Critical Regulators of Tumorigenesis, Drug Sensitivity, and Resistance. *Cancer research and treatment : official journal of Korean Cancer Association* 2021, 81(13):3431-3440.
 28. Wallis N, Oberman F, Shurrush K, ermain N, reenwald , ershon T, Pearl T, Abis , Singh V, Singh A *et al*: Small molecule inhibitor of Igf2bp1 represses Kras and a pro-oncogenic phenotype in cancer cells. *RNA biology* 2022, 19(1):26-43.
 29. Han D, Liu J, Chen C, Dong L, Liu Y, Chang R, Huang X, Liu Y, Wang J, Dougherty U *et al*: Anti-tumour immunity controlled through mRNA m(6)A methylation and YTHDF1 in dendritic cells. *Nature* 2019, 566(7743):270-274.
 30. Wan W, Ao X, Chen Q, Yu Y, Ao L, Xing W, uo W, Wu X, Pu C, Hu X *et al*: METTL3/IF2BP3 axis inhibits tumor immune surveillance by upregulating N(6)-methyladenosine modification of PD-L1 mRNA in breast cancer. *Molecular cancer* 2022, 21(1):60.
 31. Chen X, Zhou X, Wang X: m(6)A binding protein YTHDF2 in cancer. *Experimental hematology & oncology* 2022, 11(1):21.
 32. Ni H, Ruan , Sun C, Yang X, Miao Z, Li J, Chen Y, Qin H, Liu Y, Zheng L *et al*: Tanshinone IIA inhibits gastric cancer cell stemness through inducing ferroptosis. *Environ Toxicol* 2022, 37(2):192-200.
 33. Yang Y, Lu Y, Zhang C, uo Q, Zhang W, Wang T, Xia Z, Liu J, Cheng X, Xi T *et al*: Phenazine derivatives attenuate the stemness of breast cancer cells through triggering ferroptosis. *Cellular and molecular life sciences : CMLS* 2022, 79(7):360.
 34. Yuan Y, Yao H: Identification of a Novel Potent CYP4Z1 Inhibitor Attenuating the Stemness of Breast Cancer Cells through Lead Optimization. *J Med Chem* 2022, 65(23):15749-15769.
 35. Liu Y, Liang , Xu H, Dong W, Dong Z, Qiu Z, Zhang Z, Li F, Huang Y, Li Y *et al*: Tumors exploit FTO-mediated regulation of glycolytic metabolism to evade immune surveillance. *Cell metabolism* 2021, 33(6):1221-1233.e1211.
 36. Xu W, Li J, He C, Wen J, Ma H, Rong B, Diao J, Wang L, Wang J, Wu F *et al*: METTL3 regulates heterochromatin in mouse embryonic stem cells. *Nature* 2021, 591(7849):317-321.
 37. Zheng L, Xiang C, Li X, uo Q, ao L, Ni H, Xia Y, Xi T: STARD13 -correlated ceRNA network-directed inhibition on YAP/TAZ activity suppresses stemness of breast cancer via co-regulating Hippo and

- Rho-TPase/F-actin signaling. *Journal of hematology & oncology* 2018, 11(1):72.
38. Cui Q, Shi H, Ye P, Li L, Qu Q, Sun S, Sun L, Lu Z, Huang Y, Yang C *et al*: m(6)A RNA Methylation Regulates the Self-Renewal and Tumorigenesis of lioblastoma Stem Cells. *Cell reports* 2017, 18(11):2622-2634.
 39. Visvanathan A, Patil V, Abdulla S, Hoheisel JD, Somasundaram K: N⁶-Methyladenosine Landscape of lioma Stem Cell-Like Cells: METTL3 Is Essential for the Expression of Actively Transcribed genes and Sustainance of the Oncogenic Signaling. *genes* 2019, 10(2).
 40. Visvanathan A, Patil V, Arora A, Hegde AS, Arivazhagan A, Santosh V, Somasundaram K: Essential role of METTL3-mediated m(6)A modification in glioma stem-like cells maintenance and radioresistance. *Oncogene* 2018, 37(4):522-533.
 41. Chang YZ, Chai RC, Pang B, Chang X, An SY, Zhang KN, Jiang T, Wang YZ: METTL3 enhances the stability of MALAT1 with the assistance of HuR via m6A modification and activates NF-κB to promote the malignant progression of IDH-wildtype glioma. *Cancer letters* 2021, 511:36-46.
 42. Li T, Hu PS, Zuo Z, Lin JF, Li X, Wu QN, Chen ZH, Zeng ZL, Wang F, Zheng J *et al*: METTL3 facilitates tumor progression via an m(6)A-IF2BP2-dependent mechanism in colorectal carcinoma. *Molecular cancer* 2019, 18(1):112.
 43. Wang J, Dai Y, Li K, Cheng M, Xiong J, Wang X, Chen S, Chen Z, Chen J, Xu X *et al*: Deficiency of Mettl3 in Bladder Cancer Stem Cells Inhibits Bladder Cancer Progression and Angiogenesis. *Front Cell Dev Biol* 2021, 9:627706.
 44. Cao Q, Zheng J, Ni Z, Sun P, Yang C, Cheng M, Wu M, Zhang X, Yuan L, Zhang Y *et al*: The m(6)A Methylation-Regulated AFF4 Promotes Self-Renewal of Bladder Cancer Stem Cells. *Stem Cells Int* 2020, 2020:8849218.
 45. Xie J, Ba J, Zhang M, Wan Y, Jin Z, Yao Y: The m6A methyltransferase METTL3 promotes the stemness and malignant progression of breast cancer by mediating m6A modification on SOX2. *J buon* 2021, 26(2):444-449.
 46. Shi Y, Dou Y, Zhang J, Qi J, Xin Z, Zhang M, Xiao Y, Ci W: The RNA N6-Methyladenosine Methyltransferase METTL3 Promotes the Progression of Kidney Cancer via N6-Methyladenosine-Dependent Translational Enhancement of ABCD1. *Front Cell Dev Biol* 2021, 9:737498.
 47. Li H, Wang C, Lan L, Yan L, Li W, Evans I, Ruiz EJ, Su Q, Zhao J, Wu W *et al*: METTL3 promotes oxaliplatin resistance of gastric cancer CD133+ stem cells by promoting PARP1 mRNA stability. *Cellular and molecular life sciences : CMLS* 2022, 79(3):135.
 48. Weng H, Huang H, Wu H, Qin X, Zhao BS, Dong L, Shi H, Skibbe J, Shen C, Hu C *et al*: METTL14 Inhibits Hematopoietic Stem/Progenitor Differentiation and Promotes Leukemogenesis via mRNA m(6)A Modification. *Cell stem cell* 2018, 22(2):191-205.e199.
 49. Liu Z, Wu K, Wu S, Wang W, Xie S, Lu T, Li L, Dong C, Wang X, Zhou Y: A methyltransferase-like 14/miR-99a-5p/tribble 2 positive feedback circuit promotes cancer stem cell persistence and radioresistance via histone deacetylase 2-mediated epigenetic modulation in esophageal squamous cell carcinoma. *Clinical and translational medicine* 2021, 11(9):e545.
 50. Wu C, Wang Z, Zhou N, Li J, Kou Y, Luo Y, Wang Y, Yang J, Tian F: Mettl14 inhibits bladder TIC self-renewal and bladder tumorigenesis through N(6)-methyladenosine of Notch1. *Molecular cancer* 2019, 18(1):168.
 51. Wang Z, Uddin MB, Xie J, Tao H, Zeidler-Erdely PC, Kondo K, Yang C: Chronic Hexavalent Chromium Exposure Upregulates the RNA Methyltransferase METTL3 Expression to Promote Cell

- Transformation, Cancer Stem Cell-Like Property, and Tumorigenesis. *Toxicol Sci* 2022, 187(1):51-61.
52. Wang Y, Zhang L, Sun XL, Lu YC, Chen S, Pei DS, Zhang LS: NRP1 contributes to stemness and potentiates radioresistance via WTAP-mediated m6A methylation of Bcl-2 mRNA in breast cancer. *Apoptosis : an international journal on programmed cell death* 2022.
 53. Shen C, Sheng Y, Zhu AC, Robinson S, Jiang X, Dong L, Chen H, Su R, Yin Z, Li W *et al*: RNA Demethylase ALKBH5 Selectively Promotes Tumorigenesis and Cancer Stem Cell Self-Renewal in Acute Myeloid Leukemia. *Cell stem cell* 2020, 27(1):64-80 e69.
 54. Zhang S, Zhao BS, Zhou A, Lin K, Zheng S, Lu Z, Chen Y, Sulman EP, Xie K, Böglér O *et al*: m(6)A Demethylase ALKBH5 Maintains Tumorigenicity of lioblast oma Stem-like Cells by Sustaining FOXM1 Expression and Cell Proliferation Program. *Cancer cell* 2017, 31(4):591-606.e596.
 55. Kowalski-Chauvel A, Lacore M, Arnauduc F, Delmas C, Toulas C, Cohen -Jonathan-Moyal E, Seva C: The m6A RNA Demethylase ALKBH5 Promotes Radioresistance and Invasion Capability of lioma Stem Cells. *Cancers* 2020, 13(1).
 56. Zhang C, Samanta D, Lu H, Bullen JW, Zhang H, Chen I, He X, Semenza L: Hypoxia induces the breast cancer stem cell phenotype by HIF-dependent and ALKBH5-mediated m⁶A-demethylation of NANO mRNA. *Proceedings of the National Academy of Sciences of the United States of America* 2016, 113(14):E2047-2056.
 57. Yu T, Yao L, Yin H, Teng Y, Hong M, Wu Q: ALKBH5 Promotes Multiple Myeloma Tumorigenicity through inducing m(6)A-demethylation of SAV1 mRNA and Myeloma Stem Cell Phenotype. *International journal of biological sciences* 2022, 18(6):2235-2248.
 58. Liu X, Wang Z, Yang Q, Hu X, Fu Q, Zhang X, Li W: RNA Demethylase ALKBH5 Prevents Lung Cancer Progression by Regulating EMT and Stemness via Regulating p53. *Front Oncol* 2022, 12:858694.
 59. Huang H, Wang Y, Kandpal M, Zhao , Cardenas H, Ji Y, Chaparala A, Tanner EJ, Chen J, Davuluri RV *et al*: FTO-Dependent N (6)-Methyladenosine Modifications Inhibit Ovarian Cancer Stem Cell Self-Renewal by Blocking cAMP Signaling. *Cancer research* 2020, 80(16):3200-3214.
 60. Relier S, Ripoll J, uillorit H, Amalric A, Achour C, Boissière F, Vialaret J, Attina A, Debart F, Choquet A *et al*: FTO-mediated cytoplasmic m(6)A(m) demethylation adjusts stem-like properties in colorectal cancer cell. *Nature communications* 2021, 12(1):1716.
 61. Duan X, Yang L, Wang L, Liu Q, Zhang K, Liu S, Liu C, ao Q, Li L, Qin *et al*: m6A demethylase FTO promotes tumor progression via regulation of lipid metabolism in esophageal cancer. *Cell Biosci* 2022, 12(1):60.
 62. Elcheva IA, Wood T, Chiarolanzio K, Chim B, Wong M, Singh V, owda CP, Lu Q, Hafner M, Dovat S *et al*: RNA-binding protein IF2BP1 maintains leukemia stem cell properties by regulating HOXB4, MYB, and ALDH1A1. *Leukemia* 2020, 34(5):1354-1363.
 63. Myint K, Chuang LSH, Teh YX, Mawan NA, Shi EJ, Mok MMH, Nuttonmanit N, Matsuo J, Li Y, Yang H *et al*: Oncofetal protein IF2BP1 regulates IQAP3 expression to maintain stem cell potential in cancer. *iScience* 2022, 25(10):105194.
 64. Dixit D, Prager BC, imple RC, Poh HX, Wang Y, Wu Q, Qiu Z, Kidwell RL, Kim LJY, Xie Q *et al*: The RNA m6A Reader YTHDF2 Maintains Oncogene Expression and Is a Targetable Dependency in lioblastoma Stem Cells. *Cancer discovery* 2021, 11(2):480-499.
 65. Zhang C, Huang S, Zhuang H, Ruan S, Zhou Z, Huang K, Ji F, Ma Z, Hou B, He X: YTHDF2 promotes the liver cancer stem cell phenotype and cancer metastasis by regulating OCT4 expression via m6A RNA methylation. *Oncogene* 2020, 39(23):4507-4518.
 66. Hao L, Wang JM, Liu BQ, Yan J, Li C, Jiang JY, Zhao FY, Qiao HY, Wang HQ: m6A-YTHDF1-mediated

- TRIM29 upregulation facilitates the stem cell-like phenotype of cisplatin-resistant ovarian cancer cells. *Biochim Biophys Acta Mol Cell Res* 2021, 1868(1):118878.
67. Paris J, Morgan M, Campos J, Spencer J, Shmakova A, Ivanova I, Mapperley C, Lawson H, Wotherspoon DA, Sepulveda C *et al*: Targeting the RNA m(6)A Reader YTHDF2 Selectively Compromises Cancer Stem Cells in Acute Myeloid Leukemia. *Cell stem cell* 2019, 25(1):137-148.e136.
 68. Rong L, Xu Y, Zhang K, Jin L, Liu X: HNRNPA2B1 inhibited SFRP2 and activated Wnt- β /catenin via m6A-mediated miR-106b-5p processing to aggravate stemness in lung adenocarcinoma. *Pathology, research and practice* 2022, 233:153794.
 69. Chu M, Wan H, Zhang X: Requirement of splicing factor hnRNP A2B1 for tumorigenesis of melanoma stem cells. *Stem Cell Res Ther* 2021, 12(1):90.
 70. Su R, Dong L, Li Y, ao M, Han L, Wunderlich M, Deng X, Li H, Huang Y, ao L *et al*: Targeting FTO Suppresses Cancer Stem Cell Maintenance and Immune Evasion. *Cancer cell* 2020, 38(1):79-96.e11.
 71. Huang Y, Su R, Sheng Y, Dong L, Dong Z, Xu H, Ni T, Zhang ZS, Zhang T, Li C *et al*: Small-Molecule Targeting of Oncogenic FTO Demethylase in Acute Myeloid Leukemia. *Cancer cell* 2019, 35(4):677-691.e610.
 72. Huff S, Tiwari SK, onzalez M, Wang Y, Rana TM: m(6)A -RNA Demethylase FTO Inhibitors Impair Self-Renewal in lioblastoma Stem Cells. *Acs Chem Biol* 2021, 16(2):324-333.
 73. Cao K, Du Y, Bao X, Han M, Su R, Pang J, Liu S, Shi Z, Yan F, Feng S: lutathione -Bioimprinted Nanoparticles Targeting of N6-methyladenosine FTO Demethylase as a Strategy against Leukemic Stem Cells. *Small* 2022, 18(13):e2106558.
 74. Qing Y, Dong L, ao L, Li C, Li Y, Han L, Prince E, Tan B, Deng X, Wetzel C *et al*: R-2-hydroxyglutarate attenuates aerobic glycolysis in leukemia by targeting the FTO/m(6)A/PFKP/LDHB axis. *Molecular cell* 2021, 81(5):922-939.e929.
 75. Selberg S, Seli N, Kankuri E, Karelson M: Rational Design of Novel Anticancer Small-Molecule RNA m6A Demethylase ALKBH5 Inhibitors. *ACS omega* 2021, 6(20):13310-13320.
 76. Yankova E, Blackaby W, Albertella M, Rak J, De Braekeleer E, Tsagkogeorga , Pilka ES, Aspris D, Leggate D, Hendrick A *et al*: Small-molecule inhibition of METTL3 as a strategy against myeloid leukaemia. *Nature* 2021, 593(7860):597-601.
 77. Xie , Wu XN, Ling Y, Rui Y, Wu D, Zhou J, Li J, Lin S, Peng Q, Li Z *et al*: A novel inhibitor of N (6)-methyladenosine demethylase FTO induces mRNA methylation and shows anti-cancer activities. *Acta pharmaceutica Sinica B* 2022, 12(2):853-866.
 78. Feng S, Qiu , Yang L, Feng L, Fan X, Ren F, Huang K, Chen Y: Omeprazole improves chemosensitivity of gastric cancer cells by m6A demethylase FTO-mediated activation of mTORC1 and DDIT3 up-regulation. *Bioscience reports* 2021, 41(1).
 79. Sun K, Du Y, Hou Y, Zhao M, Li J, Du Y, Zhang L, Chen C, Yang H, Yan F *et al*: Saikosaponin D exhibits anti-leukemic activity by targeting FTO/m(6)A signaling. *Theranostics* 2021, 11(12):5831-5846.
 80. Chen WW, Qi JW, Hang Y, Wu JX, Zhou XX, Chen JZ, Wang J, Wang HH: Simvastatin is beneficial to lung cancer progression by inducing METTL3-induced m6A modification on EZH2 mRNA. *European review for medical and pharmacological sciences* 2020, 24(8):4263-4270.
 81. Zhao Z, Zeng J, uo Q, Pu K, Yang Y, Chen N, Zhang , Zhao M, Zheng Q, Tang J *et al*: Berberine Suppresses Stemness and Tumorigenicity of Colorectal Cancer Stem-Like Cells by Inhibiting m(6)A Methylation. *Front Oncol* 2021, 11:775418.